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Multi-parametric neuroimaging reproducibility: A 3-T resource study

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ABSTRACT

Modern MRI image processing methods have yielded quantitative, morphometric, functional, and structural assessments of the human brain. These analyses typically exploit carefully optimized protocols for specific imaging targets. Algorithm investigators have several excellent public data resources to use to test, develop, and optimize their methods. Recently, there has been an increasing focus on combining MRI protocols in multi-parametric studies. Notably, these have included innovative approaches for fusing connectivity inferences with functional and/or anatomical characterizations. Yet, validation of the reproducibility of these interesting and novel methods has been severely hampered by the limited availability of appropriate multiparametric data. We present an imaging protocol optimized to include state-of-the-art assessment of brain function, structure, micro-architecture, and quantitative parameters within a clinically feasible 60-min protocol on a 3-T MRI scanner. We present scan–rescan reproducibility of these imaging contrasts based on 21 healthy volunteers (11 M/10 F, 22–61 years old). The cortical gray matter, cortical white matter, ventricular cerebrospinal fluid, thalamus, putamen, caudate, cerebellar gray matter, cerebellar white matter, and brainstem were identified with mean volume-wise reproducibility of 3.5%. We tabulate the mean intensity, variability, and reproducibility of each contrast in a region of interest approach, which is essential for prospective study planning and retrospective power analysis considerations. Anatomy was highly consistent on structural acquisition (~1-5% variability), while variation on diffusion and several other quantitative scans was higher (~<10%). Some sequences are particularly variable in specific structures (ASL exhibited variation of 28% in the cerebral white matter) or in thin structures (quantitative T2 varied by up to 73% in the caudate) due, in large part, to variability in automated ROI placement. The richness of the joint distribution of intensities across imaging methods can be best assessed within the context of a particular analysis approach as opposed to a summary table. As such, all imaging data and analysis routines have been made publicly and freely available. This effort provides the neuroimaging community with a resource for optimization of algorithms that exploit the diversity of modern MRI modalities. Additionally, it establishes a baseline for continuing development and optimization of multi-parametric imaging protocols.

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Introduction

Magnetic resonance imaging (MRI) enables non-invasive study of the structure, functional response, and connectivity of the *in vivo* human nervous system with millimeter resolution. Visual assessment of these MRI data by radiologists has become an extraordinarily important part of patient care in terms of improving diagnosis, prognosis, and monitoring, especially for neurological disease. Image

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processing methods have yielded quantitative, morphometric, functional, and structural assessments of the human brain. These analyses typically exploit carefully optimized protocols for specific imaging targets, so that, historically, a particular algorithm extracts a piece of information from a particular kind of image. There are several excellent public data resources that algorithm investigators may use to test, develop, and optimize their methods. Recently, there has been an increasing focus on combining MRI protocols in multi-parametric studies, most notably leading to the current emphasis on the study of the physical and functional connections between different brain regions (i.e., the "connectome" (Sporns et al., 2005)). These have included innovative approaches for fusing connectivity inferences

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with functional and/or anatomical characterizations. Yet validation of the reproducibility of these interesting and novel methods has been severely hampered by the limited availability of appropriate multiparametric data.

MRI reproducibility is commonly assessed with protocols optimized to answer specific questions. However, scan time is a precious resource. Construction of a general protocol that covers many different imaging sequences to meet a specific time constraint (herein, 1 h) leads to comprises based on experience and "arbitrary" judgments. The aggregate time constraint certainly limits the quality of data that can be acquired, especially for sequences that require a relatively large amount of data (e.g., parametric mapping for relaxometry, diffusion, etc.), but time is a real constraint under which most clinical studies must labor. MRI data are more often being reused for purposes other than that for which originally intended, so it may be pragmatic to use a more general imaging sequence that leave open greater possibilities for reanalysis. Herein, we present and characterize one such unified sequence for multi-parametric MRI.

Historically, data resources have been made available in the form of published atlases, with early and lasting contributions from Brodmann (1907) and Talairach and Tournoux (1988). These stereotaxic spaces have been digitized and widely applied to 3D MRI interpretation. The Montreal Neurological Institute (MNI) and the International Consortium for Brain Mapping (ICBM), in particular, have released a series of brain atlases (including the large-scale averages MNI305, MNI152, and MNI452 along with the high-resolution "Colin" brain), which have come to define standard stereotaxic space in MRI-based neuroscience. In line with the NIH data-sharing intuitive, full datasets have begun to emerge; these enable substantively more general re-analyses than are possible with average or representative atlases. Two prominent initiatives, the Alzheimer's Disease Neuroimaging Initiative (ADNI) (Mueller et al., 2005) and Open Access Series of Imaging Studies (OASIS), have provided a substantial number of public MR human brain datasets. Although large in their participant numbers, both of the datasets are limited in the cataloging of various MR modalities. The ADNI project consists of approximately 819 subjects scanned longitudinally at both 1.5 and 3 T with T1, T2, and proton density (PD) weighted sequences (Jack et al., 2008; Mueller et al., 2005). The OASIS project is divided into longitudinal (Marcus et al., 2010) and cross-sectional (Marcus et al., 2007) sub-studies with T2-weighted images. The cross-sectional dataset consists of 416 subjects (18-96 years old). The longitudinal dataset contains 150 subjects (60-96 years old). Numerous smaller datasets are available through the Biomedical Informatics Research Network (BIRN, http://www.birncommunity.org/resources/data/) and various investigators' websites. While some particularly exciting datasets are available (for example, the "1000 Functional Connectomes" Project—http://www.nitrc.org/projects/fcon_1000/), no substantial scan-rescan datasets with a broad range of multi-parametric MRI studies have been publicly disseminated (to the authors' knowledge).

In other imaging disciplines, richly featured, common resources libraries have spurred innovation of both general and specialized methods (consider the Berkeley Segmentation Dataset and Benchmark-http://www.eecs.berkeley.edu/Research/Projects/CS/vision/ bsds/). The lack of widely available data appropriate for validation of image processing algorithms that make use of multi-parametric MRI data has hampered the development and validation of such algorithms. Herein, we present an imaging protocol optimized to include state-of-the-art assessment of brain function, structural characteristics, micro-architecture, and quantitative parameters within a clinically feasible 60-min protocol at 3 T. We assess scanrescan reproducibility of these imaging contrasts and report the results in a tabular form for convenient prospective study planning and retrospective power analysis considerations. The richness of the joint distribution of intensities across imaging methods can be best assessed within the context of a particular analysis approach as opposed to a summary table. As such, all imaging data and analysis routines have been made publicly and freely available. This effort provides the neuroimaging community with a resource for optimization of algorithms that may exploit the diversity of modern MRI modalities and establishes a baseline for continuing development and optimization of multi-parametric imaging protocols.

Methods

Twenty-one healthy volunteers with no history of neurological conditions (11 M/10 F, 22-61 years old) were recruited. Local institutional review board approval and written informed consent were obtained prior to examination. All data were acquired using a 3-T MR scanner (Achieva, Philips Healthcare, Best, The Netherlands) with body coil excitation and an eight channel phased array SENSitivity Encoding (SENSE) (Pruessmann et al., 1999) head-coil for reception. Subjects were scanned twice with one of two protocols listed in Table 1 (either the short protocol or the extended protocol). The longer protocol fully used the allotted scan time, while the shorter protocol allowed for a less rushed transition between scans. The protocol used for each subject was selected based on scanner availability. Ten subjects were scanned with the short protocol and 11 subjects with the long protocol. After being scanned for approximately 1 h, each subject exited the scan room for a short break and then reentered the scan room for an identical session. Notably, there was a full repositioning of the volunteer, coils, blankets, and pads before both 1-h sessions. All scans were completed during a 2-week interval. The resulting dataset consisted of 42 "1-h" sessions of 21 individuals. Randomized ID codes (01... 42) were assigned to each of the sessions, and each individual was assigned a unique 3-digit key. The sequences explored for this study are briefly reviewed in the following sections.

To assess the intrinsic SNR of the sequences in the absence of repositioning effects, a single subject (M/25 years old) was scanned twice with the long protocol without repositioning between sessions. This scan took place 9 months after the original scan session.

Data

Structural imaging

Conventional MR imaging has been used to elicit contrast between neighboring tissues, between tissues types, and between tissues that are diseased and healthy. The two "conventional" sequences that were used in this study – FLAIR and MPRAGE, as detailed next – take advantage of inversion recovery pulses and the fact that each tissue has a characteristic longitudinal relaxation (T1).

FLAIR

FLuid Attenuated Inversion Recovery (FLAIR) has been in service since 1992 (Hajnal et al., 1992) and has been perhaps the most commonly used imaging sequence for detecting and diagnosing diseases of the central nervous system (Barkhof and Scheltens, 2002; Kastrup et al., 2005; Rumboldt and Marotti, 2003). FLAIR sequences exploit the idea that when signal from the cerebrospinal fluid (CSF) is nulled, T2 hyperintensities of the brain tissue will reflect lesions better (Hajnal et al., 1992).

FLAIR description

FLAIR MRI is performed by applying an inversion pulse prior to a spin-echo (or fast spin-echo) readout and can be performed in a multi-slice or a 3D fashion. The 3D FLAIR sequence (so-called VISTA by Philips Medical Systems and SPACE by Siemens Medical Systems) is a multi-echo-turbo-spin-echo, with select modifications: (1) the refocusing pulse train is modulated in such a fashion to maintain a very high SNR and CNR scan throughout an extended readout train. Busse et al. noted that with appropriate RF modulation of the

Table 1 Multi-parametric brain assessment protocol definition.

Sequence	Target	Orientation	Scan time		Acquired resolution (mm)			
		(type) ^a	Minutes	Seconds	Left-right	AntPos.	Foot-head	Sense
Triplanar survey	=	Localizer	0	32	250	250	50	-
Reference scan	_	Reference	0	45	450	300	300	_
FLAIR	Structural	Sag (3D)	8	48	1.1	1.1	1.1	2×2
MPRAGE (Fast ADNI)	Structural	Sag (3D)	5	56	1	1	1.2	2
Diffusion tensor imaging (DTI)	Diffusion	Axi (MS)	4	11	2.2	2.2	2.2	2.5
DTI geometric reference	Diffusion	Sag (3D)	1	48	2	2	2	2×2
Resting state fMRI	Functional	Axi (MS)	7	14	3	3	3 ^b	2
B_0 field map (TE = 8 ms)	Field map	Axi (MS)	0	40	2.4	2.4	3	2
B_0 field map (TE = 9 ms)	Field map	Axi (MS)	0	40	2.4	2.4	3	2
B_1 field map ^c	Field map	Axi (MS)	2	45	4	4	4	2
Arterial spin labeling (ASL)	Perfusion	Axi (MS)	4	56	3	3	7 ^b	2.5
Arterial spin labeling (M_0)	Perfusion	Axi (MS)	0	48	3	3	7 ^b	2.5
VASO	Perfusion	Axi (3D)	3	45	3	3	5	3
qT1 (15°) ^c	Quantitative	Axi (3D)	3	40	0.83	0.83	3	1
qT1 (60°) ^c	Quantitative	Axi (3D)	3	40	0.83	0.83	3	1
qT2 (dual echo)	Quantitative	Axi (MS)	4	13	1.5	1.5	1.5	2
Magnetization transfer	Quantitative	Axi (3D)	3	09	1.5	1.5	1.5	2.5

- ^a Sag: Sagittal. Axi: Axial. 3D: Three-dimensional Fourier acquisition. MS: Multi-slice acquisition.
- ^b Sequence includes a 1 mm slice gap.
- ^c Sequence was not included in the "short" 1 h protocol.

refocusing pulse train, one can maintain the echo for a very long time while creating the contrast from the earlier echoes (Busse, 2004; Busse et al., 2006; Chagla et al., 2008). (2) We image in the sagittal plane such that SENSE is applied in 2 directions, phase encode in 2 directions and turbo directions in 2 directions (y and radial). Hence, the sampling of k-space is not a simple linear sampling pattern therefore when coupled with modulations of RF refocusing; this yields the desired T2-contrast and high CNR over long TE. Furthermore, Busse et al. noted that due to the RF modulation, the T2 decay is not a single exponential with decay constant = T2. It is, however, decaying with some other decay constant (much longer than T2). The reason being is that if the magnetization is not fully in the x-y plane, then it will dephase less rapidly. Because the apparent T2 is much longer, the extended echo train does not blur the image (T2-filter, or "smearing") in quite the same way as a conventional 90-180-180-180..., turbo-spin-echo (TSE) sequence.

The delay time between the inversion pulse and the excitation of the slice (or phase encode for 3D) is approximately the time when the CSF signal crosses the zero-point on the T1 recovery curve (Hajnal et al., 1992). The timing for CSF nulling is proportional to the T1 of CSF, so the inversion time (TI) for FLAIR sequences is often very long (TI~2400 ms), and the TR between excitations is longer still (~10 s) to allow for full recovery of the longitudinal magnetization of the CSF prior to the subsequent inversion. To shorten the scan time and gain an increase in resolution, 3D methods have been employed as in this article (Geurts et al., 2008; Moraal et al., 2008; Yoshida et al., 2008). The sequence parameters for the FLAIR scan for this study are as follows. A 3D inversion recovery sequence was played out (TR/TE/TI = 8000/330/2400 ms) with a $1.1 \times 1.1 \times 1.1 \text{ mm}^3$ resolution over an FOV of $242 \times 180 \times 200 \text{ mm}$ (anterior–posterior, right–left, foot–head) acquired in the sagittal plane. The SENSE acceleration factor was 2 in the right-left direction. Multishot, RF amplitude-modulated fast spin-echo (TSE factor = 110) was used with 6 dummy start-up echoes. To reduce the power deposition resulting from a large number of refocusing pulses, minimize the effect of T2-decay over a long TE, and reduce the T2-filtering effect, the refocusing angle was set to 50°, although the actual flip angle of refocusing pulses is modulated similarly to the methods presented by Busse et al. Fat saturation was applied using an adiabatic inversion recovery (SPAIR, TI = 220 ms). The total scan time was 8 min 48 s.

MPRAGE

MPRAGE (Magnetization-Prepared Rapid Acquisition Gradient Echo) (Mugler and Brookeman, 1990) uses an inversion pulse played

out prior to the imaging sequence and timed such that the signal from gray matter (GM) is very low compared to the surrounding white matter (WM). The result is a high-resolution, high-contrast T1-weighted imaging method that can be used to segment the WM from the GM and CSF. Neither FLAIR nor MPRAGE acquisitions are quantitative in nature—i.e., the observed intensities do not correspond to a specific biophysical parameter. Quantitative analyses of these images rely on morphometrics or tissue contrast modeling.

MPRAGE sequence

MPRAGE acquisitions employ a gradient echo readout with a very short TE, the use of which limits the susceptibility-related artifacts that occur at the air–tissue interfaces. The sequence parameters for the MPRAGE scan for this study are as follows. A 3D inversion recovery sequence was played out (TR/TE/TI = 6.7/3.1/842 ms) with a $1.0\times1.0\times1.2$ mm³ resolution over an FOV of $240\times204\times256$ mm acquired in the sagittal plane. The SENSE acceleration factor was 2 in the right–left direction. Multi-shot fast gradient echo (TFE factor=240) was used with a 3-s shot interval and the turbo direction being in the slice direction (right–left). The flip angle=8°. No fat saturation was employed. The total scan time was 5 min 56 s.

Diffusion weighted imaging

Diffusion weighted MRI (DWI) measures the spatial diffusion characteristics of water molecules and can provide information about the microstructural environment within a voxel (Beaulieu, 2002).

Diffusion tensor MRI

Following early demonstration that DWI is highly sensitive to pathological changes in acute stroke (Miyabe et al., 1996; Moseley et al., 1990a,b; Ulug et al., 1997; van Gelderen et al., 1994), a key development was the introduction of diffusion tensor formalism (Basser et al., 1994a,b; van Zijl et al., 1994), which forms the basis of diffusion tensor imaging (DTI). DTI has been widely used to establish the connectivity of WM fiber tracts using the anisotropy and principal orientation of diffusion in each voxel (Mori et al., 1998), which allow for fiber tracking (Mori and van Zijl, 2002).

DTI sequence

The DTI dataset was acquired with a multi-slice, single-shot, echoplanar imaging (EPI), spin-echo sequence (TR/TE = 6281/67 ms, SENSE factor = 2.5). Sixty-five transverse slices were acquired parallel

to the line connecting the anterior commissure (AC) to the posterior commissure (PC) with no slice gap and 2.2 mm nominal isotropic resolution (FOV = 212 × 212, data matrix = 96 × 96, reconstructed to 256 × 256). Fat suppression was performed with Spectral Presaturation with Inversion Recovery (SPIR) and the phase encoding direction was anterior–posterior. Diffusion weighting was applied along 32 directions (Philips parameters: gradient overplus = no, directional resolution = high, gradient mode = enhanced) with a *b*-value of 700 s/mm². The amplitude, leading edge spacing, and duration of the magnetic field gradients were G=49 mT/m, $\Delta=32.3$ ms, and $\delta=12$ ms, respectively). The diffusion time, $t_{\rm dif}$, was 28.3 ms ($t_{\rm dif}=\Delta-\delta/3$). Five minimally weighted images (5 B_0 with $b\approx33$ s/mm²) were acquired and averaged on the scanner as part of each DTI dataset. The total scan time to acquire the DTI dataset was 4 min 11 s. No cardiac or respiratory gating was employed.

To provide a reference dataset for image-based distortion correction of DTI data (e.g., the "DTI Geometric Reference" scan), a T2-weighted volume with minimal distortion was acquired with the same planning geometry using a 3D multi-shot turbo spin–echo with a TSE factor of 100 (linear profile order, 6 startup echoes, 3D non-selective 90° excitation pulse followed by one 180° refocusing pulse and a train of 99 35° refocusing pulses, TR/TE = 2500/287 ms, SENSE factor 2 in the anterior–posterior and right–left directions). 180 Sagittal slices were acquired with a nominal resolution of 2.0×2.0 mm in–plane and a slice thickness of 1 mm (FOV = $200 \times 242 \times 180$ mm, acquired matrix 100×118 , reconstructed matrix 256×256). Slices were overlapping with a resampling factor of 1. Fat suppression as performed with SPIR and the phase encoding direction was anterior–posterior. Two signal averages were performed (NSA = 2), and the total scan duration was 1 min 47 s.

Functional imaging

Resting state functional MRI (fMRI)

Functional connectivity (FC) in the brain refers to temporally synchronous neural activity from spatially distinct locations. Biswal et al. (1995) introduced FC in blood oxygen level-dependent (BOLD) fMRI by investigating the connectivity of a region within the motor cortex during "rest". In recent years, BOLD fMRI has been widely used to evaluate FC revealing several neurobiologically relevant intrinsically connected networks (ICNs) (Greicius et al., 2003; Raichle et al., 2001; Seeley et al., 2007; Vincent et al., 2004). Note that physiological processes (respiration rate, depth of respiration, cardiac pulsation) also induce low-frequency temporally correlated signals from distant (often bilateral) regions in the brain (Behzadi et al., 2007; Glover et al., 2000) should be used while investigating FC.

fMRI sequence

The sequence used for resting state functional connectivity MRI is typically identical to that used for BOLD functional MRI studies of task activation. Here, we used a 2D EPI sequence with SENSE partial-parallel imaging acceleration to obtain 3×3 mm (80 by 80 voxels) inplane resolution in thirty-seven 3 mm transverse slices with 1 mm slice gap. An ascending slice order with TR/TE = 2000/30 ms, flip angle of 75°, and SENSE acceleration factor of 2 were used. SPIR was used for fat suppression. This study used an ascending slice acquisition order because a pilot studies revealed smaller motion induced artifacts with ascending slice order than with interleaved slice order. While using an ascending slice order, it was necessary to use a small slice gap to prevent cross talk between the slices. One 7-min run was recorded which provided 210 time points (discarding the first four volumes to achieve steady state).

One of the primary sources of artifacts in functional connectivity is due to correlation of signals from non-neuronal physiological activity such as respiration and cardiac pulsation. To minimize these effects, several methods have been proposed. One such method is RETROICOR (Glover et al., 2000), which uses respiration signal (typically from a

respiratory belt) and a cardiac pulse signal (typically from a pulse oximeter). A newer method (COMPCOR) uses signal from the white matter and CSF to minimize similar artifacts and has been shown to be equally good if not better than RETROICOR (Behzadi et al., 2007). In light of these computational methods, we did not record respiratory or cardiac signals and suggest using COMPCOR to minimize physiology induced artifacts.

Field mapping

B₀ mapping

MRI field maps provide information about the local magnetic field inhomogeneity, which is an important contributing factor for artifacts and signal loss (Bankman, 2008). To study this field, we acquire a phase map using a gradient echo sequence for two different echo times (Haacke, 1999; Rajan, 1998; Schneider and Glover, 1991; Webb and Macovski, 1991). Subtraction of the two phase maps generates an image from which the field inhomogeneity can be computed (Andersson et al., 2003; Cusack et al., 2003; Gholipour et al., 2008; Reber et al., 1998).

*B*₀ *fieldmap sequence*

Two sequential 2D, gradient echo multi-slice volumes were acquired with differing echo times (8 ms versus 9 ms). The remaining scan parameters were identical for the acquisitions (TR=745 ms, 3 mm slice thickness, 92×92 acquisition matrix, $220\times220\times150$ mm FOV). Total scan time was 1 min 20 s.

B_1 mapping

Excitation of the magnetization in a sample is caused by the active transverse component of the transmitted radiofrequency (RF) field, which varies over spatial location (Stollberger and Wach, 1996). Nonuniformities in the RF field contribute to image variation between different experiments; these variations stem from the RF coil itself, the dielectric resonance of the sample, the shape of the sample, induced RF eddy and displacement currents, disparities between machinery, and many other variables (Samson et al., 2006; Stollberger and Wach, 1996). The spatial distribution of the transmitted RF field can affect quantitative measurements, including the assessment of the longitudinal relaxation time, magnetization transfer ratio, specific absorption rate (SAR) calculations, etc. Several methods for B_1 mapping have been proposed, including determining the voltage induced in a small pick-up coil within the larger transmit coil (Hornak et al., 1988), fitting signal equations for serial scans with variable pulse powers or durations (Hornak et al., 1988), or utilizing the ratio between two images to compare variances in signal according to changing flip angles (Insko and Bolinger, 1993; Rudolf and Paul, 1996), or changing repetition times (Yarnykh, 2007).

B_1 sequence

Here, we use the Actual Flip-Angle Imaging (AFI) to produces B_1 maps using a single-scan pulsed steady-state acquisition, which is insensitive to T1, T2, T2*, spin density, coil reception profile, and flow or motion artifacts (Yarnykh, 2007). The AFI method consists of two identical pulses with the same flip angle applied before two different delay times (TR₁ and TR₂). This sequence works under the assumptions that $TR_1 < TR_2 < T1$, so that the fast repetition rate of the sequence establishes a pulsed steady state of magnetization and that the sequence is ideally spoiled so that all transverse coherences are destroyed at the end of TR₁ and TR₂. The optimal range of TR₁ is between 10 and 30 ms, a flip angle between 40° and 80°, and a ratio of TR₂/TR₁ between 4 and 6, which usually results in a 15–65% decrease of S_2 from S_1 (Yarnykh, 2007). In this study, we acquired a B_1 map with a 3D gradient echo sequence using the in-scanner image intensity equalizing algorithm (i.e., Constant Level AppeaRance, CLEAR) and 3D spoiled gradient echo sequence. On-scanner calculations produced a B_1 map as the ratio of signals obtained after each of the two slice

selective excitation pulses relative to the nominal flip angle (alpha = 60°) consisting of 43 axial slices. Other imaging parameters were TR₁/TR₂/TE/alpha = 12 ms/60 ms/2.3 ms/ 60° , gradient spoil factor = 10, nominal in-plane resolution = 4 mm × 4 mm, slice thickness = 4 mm, FOV = $220 \times 220 \times 172$ mm. The total scan duration was 2 min 45 s.

Imaging of perfusion (blood flow and blood volume)

Arterial spin labeling (ASL)

Cerebral perfusion, defined as the rate that blood is delivered to brain tissue, can be measured with dynamic susceptibility contrast (DSC-MRI) using gadolinium chelates. Arterial spin labeling (ASL) was developed as a non-invasive alternative (Detre et al., 1992; Williams et al., 1992) in the respect that it does not require injection of any contrast agent because blood water as an endogenous tracer. In ASL, two images are acquired: a "labeled" image where blood water is inverted in the feeding arteries using RF pulses and a "control" image where static tissue signals are identical. After labeling the blood and waiting a period of time for the labeled blood to exchange with the tissue (Alsop, 1997; Buxton, 1993; Golay et al., 2002), images are acquired. The difference of the labeled and control image yields an image that is proportional to the cerebral blood flow.

ASL sequence

ASL images were acquired using a single-shot gradient echo EPI (SENSE = 2.5, label duration = 1650 ms, TR/TE/TI = 4000/14 ms/1525).The balanced pseudocontinuous labeling scheme was used with maximum/mean gradient = 6/0.6 mT/m (Edelman et al., 1994; van Osch et al., 2009). Other scan parameters include $FOV = 240 \times 240$ mm, voxel size $=3\times3\times7$ mm, 19 slices, 35 pairs of controls/labels, RF interval 1 ms, RF duration 0.5 ms, and flip angle 18°. Labeling efficiency was estimated to be 0.85 based on simulations by Wu et al. (2007). The imaging slices were centered along the AC-PC, and labeling occurred 84 mm below the AC-PC line. To calculate the magnetization of arterial blood (M_0) , an inversion recovery sequence with the same geometry and resolution as the ASL sequence (inversion times 100-1900 ms with 200-ms intervals) was acquired. M_0 was quantified using the same assumptions and according to the same procedure outlined by Chalela et al. (2000) (van Osch et al., 2009). Scan time was 4 min 56 s for the brain ASL sequence and 48 s for the M_0 reference. Perfusion was not quantified.

Vascular space occupancy (VASO)

Cerebral blood volume (CBV) changes are essential for understanding the BOLD signal mechanism and can be used as an intrinsic contrast for fMRI. Conventionally, CBV can be determined using computer tomography (CT) and PET (Hoeffner et al., 2004). More recently, a few MRI techniques have been developed to measure CBV *in vivo* without ionizing radiation (Kuppusamy et al., 1996; Moseley et al., 1992; Ostergaard et al., 1996a,b; Schwarzbauer et al., 1993). Vascular space occupancy (VASO) MRI is an approach that is sensitized to CBV changes in the brain without the need for contrast agent injection (Lu et al., 2003).

VASO sequence

The VASO sequence is an inversion recovery-based approach. The blood nulling time is determined by the field (B_0) -dependent blood T1 relaxation time (Lu et al., 2004) and user-defined scan repetition time (TR). A spatially non-selective adiabatic inversion pulse is commonly used, which is followed by imaging modules such as gradient echo EPI, spin echo EPI, and turbo-spin-echo (TSE). Recent studies (Donahue et al., 2009a; Poser and Norris, 2009) have demonstrated that the gradient spin-echo (GraSE) sequence may be a better choice for VASO due to improved SNR and efficiency. In this study, we employed a 3D GraSE sequence. Imaging parameters were TR/TE/TI/FA = 5000 ms/

20 ms/1054 ms/90°, TSE/EPI direction = foot-head/anterior-posterior, TSE/EPI factor = 15/13, imaging matrix = 64×64 , in-plane resolution = 3×3 mm, slice thickness = 5 mm, 25 slices, SENSE = 2. To minimize the contamination from the BOLD effect, short TE is preferred in VASO sequences; it has been shown that the image contrast in VASO sequences with long TR (>5 s) is dominated by CBV change, whereas short-TR (<2 s) VASO signal has contributions from side effects such as CBF, fresh inflow blood and partial-volume effects from WM and CSF (Donahue et al., 2009b; Donahue et al., 2006). Total scan time was 3 min 45 s.

Quantitative relaxometry imaging

T1 mapping

Understanding the T1 of tissues in the human body has been paramount in devising sequences that optimize contrast for clinically utilized sequences such as FLAIR (Hajnal et al., 1992) and MPRAGE (Mugler and Brookeman, 1990), and developing quantitative T1-based methods, such as ASL (Detre et al., 1992; Williams et al., 1992), vascular space occupancy (Lu et al., 2003). By definition, quantitative T1 imaging is the assessment of the absolute T1 relaxation time in various tissues.

T1 mapping sequence

T1 can be mapped by a series of scans are performed at different T1 values, but, although a robust method, this method is hindered by extremely long examination times (\sim minutes/slice), susceptibility to motion, and exchange between the intracellular and extracellular spaces in the long inversion time regime. Here, we use a multi-flip angle (two or more) whereby a series of conventional gradient echo images are obtained using different excitation flip angles (Bottomley and Ouwerkerk, 1994). The difficulties with this method hinge on the fact that it is expected that the excitation flip angles are performed precisely as prescribed. Therefore, any B_1 inhomogeneity can confound the measurement of T1 by as much as 30% in the human brain (Yarnykh, 2007). To overcome this shortcoming, a B_1 map can be obtained and the T1 values corrected accordingly.

Generally the flip angles chosen are fairly low to encourage better B_1 performance (Bottomley and Ouwerkerk, 1994); here, we chose flip angles = 15° and 60° with a spoiled 3D gradient echo with FOV = 212 × 212 × 143 mm and acquired resolution = 1.5 × 1.5 × 1.5 mm. SENSE acceleration factor = 2.5 with multi-shot EPI (factor = 7). TR/TE/ α 1/ α 2 = 100 ms/15 ms/15°/60°. Two k-space averages were performed, and the total scan time was 3 min 40 s per flip angle (7 min 20 s total).

T2 mapping

Understanding the T2 of various tissues in the human body has been critical in optimizing contrast in clinical sequences and T2weighted imaging is one of the most prolific markers for detecting and following lesions and their evolution in neurodegenerative diseases (Lee et al., 1998; Lu et al., 2005b). Two methods have been employed to measure the absolute T2 of tissue. The first is the multiple echo method, where a spin-echo sequence is performed, and the signal as a function of echo time is obtained, but, as with T1 mapping, this technique is prohibitively long with some acquisitions lasting over 30 min. To overcome these hindrances, T2 can be obtained from a twopoint method, where one dataset is obtained with long TR and short TE (i.e., proton-density weighting) and a second obtained with the same TR and a long TE (~T2, T2-weighted). The advantage of this method is that in a typical clinical evaluation, PD- and T2-weighted images are already obtained, thus the calculation of the estimate of T2 follows with little effort.

T2 mapping sequence

The double echo time T2 measurement is a conventional spinecho with data obtained at two distinct echo times. It is important to keep in mind that the TR should be long enough to allow for sufficient relaxation prior to the next excitation. In order to shorten the scan time even further, often fast readout trains (fast spin–echo, FSE) are used. Post hoc, a T2 map is obtained. The sequence that we performed is a multi-slice spin–echo with FOV = $212 \times 212 \times 143$ mm and acquired resolution = $1.5 \times 1.5 \times 1.5$ mm. SENSE acceleration factor = 2.0 with multi-shot fast spin–echo (TSE, TSE factor = 8). TR/TE1/TE2 = 6653 ms/ 30 ms/80 ms. The total scan time was 4 min 12 s.

Magnetization transfer (MT) imaging

MT imaging is exquisitely sensitive to the macromolecular composition of tissue, and particularly in the central nervous system, alterations in the myelin content (Schmierer et al., 2004). Conventionally, two types of MT imaging are used in human studies. Continuous wave (CW) (Henkelman et al., 2001; Stanisz et al., 2005; Wolff and Balaban, 1989) experiments rely on the large saturation effects induced by a single, long, continuous (i.e., constant amplitude) RF irradiation within a single TR period, while pulsed MT imaging (Sled and Pike, 2000; Yeung and Aisen, 1992) employs a series of short (~10-30 ms), high-amplitude RF saturation pulses repeated over multiple, short-TR periods. Pulsed MT affords much greater acquisition speed and has provided the ability to survey the frequency dependence of the MT effect and to determine MT-based parameters such as fractional pool sizes and exchange rates (Portnoy and Stanisz, 2007; Sled and Pike, 2001; Yarnykh and Yuan, 2004; Smith et al., 2006).

MT sequence

A pulsed MT sequence was performed with a spoiled 3D gradient echo with FOV = $212 \times 212 \times 143$ mm and acquired resolution = $1.5 \times 1.5 \times 1.5$ mm. SENSE acceleration factor = 2.5 with multi-shot EPI (factor = 9). TR/TE/ α = 64 ms/15 ms/9°. MT preparation was achieved using a five-lobed, sinc-gauss shaped RF irradiation (B_1 = 10.5mT, duration = 24 ms, and offset frequency = 1500 Hz). The reference scan was obtained with the same parameters in the absence of an MT preparation. Two k-space averages were performed, and the total scan time was 3 min 9 s.

Analysis

All imaging data were acquired in PAR/REC format (Philips Healthcare, Best, The Netherlands) and converted to Neuroimaging Informatics Technology Initiative (NifTI) format, Names, dates, and text fields were removed from PAR files for anonymity. Randomized codes were assigned to each subject ("Kirby###") and to each scan session (01... 42). Anonymized NIfTI data were analyzed in a subject-specific space, which was defined in the following manner. One MPRAGE for each subject was randomly selected as the target for each subject (21 MPRAGE images), and the remaining 21 MPRAGE images were registered with a six-degree-of-freedom rigid body transform (with cubic spline interpolation) into this space, resulting in a registered MPRAGE for each scan session. The fMRI and DTI scans were registered to the coregistered T2w anatomical target (which is not susceptible to EPI distortion) of their own session using affine (12-degree-of-freedom) registration to compensate for EPI distortion. Other scans within the "first" session were registered to the "first" session MPRAGE, while all other scans within the "second" session were registered to the transformed MPRAGE from the second sessions using rigid body registration and normalized mutual information criteria.

The brain was isolated with SPECTRE, a joint registration-segmentation approach (Carass et al., 2007) to form "skull-stripped" MPRAGE. TOpology-preserving, Anatomy-Driven Segmentation (TOADS) (Bazin and Pham, 2007) was used to label the cerebral gray matter, cerebral white matter, cerebellar gray matter, cerebellar white matter, brainstem, brainstem, caudate, putamen, thalamus, and ventricular cerebrospinal fluid (CSF). For each of the labeled regions, the following were assessed in a cross-sectional manner: (1) the

mean regional intensity, (2) the coefficient of variation of the intensity within the region, (3) the coefficient of variation of intensity between scans, and (4) the intra-class coefficient (ICC). The ICC assesses the degree of absolute agreement among measurements (i.e., criterion-referenced reliability—also known as A-1) (McGraw and Wong, 1996; Shrout and Fleiss, 1979). For scans with multiple modalities (e.g., the 32 diffusion weighted directions used in DTI), we report the average coefficient of variation and ICC over the series of scan types.

To assess SNR, three regions were manually delineated in the corpus callosum, peripheral white matter, and the globus palidus using the MPRAGE image. SNR was calculated on the scan–rescan data without repositioning as the $\sqrt{2}$ times the mean signal intensity divided by the standard deviation of the scan–rescan difference within each region.

All data analysis was performed using the Java Imaging Science Toolkit (JIST) (Lucas et al., 2010), an open-source framework for medical image analysis and batch processing based on the Medical Image Processing, Analysis, and Visualization (MIPAV) program produced by the Center for Information Technology, National Institutes of Health (McAuliffe et al., 2001). Binaries and source code are available through the Neuroimaging Informatics Tools and Resources Clearinghouse (NITRC) resource website (http://www.nitrc.org/projects/jist/). Summary statistics were computed with MATLAB (The Mathworks, Natick, MA).

There is a rich and diverse body of literature on each imaging modalities employed in this study. In this section, we provide a brief indication of how the data are "typically" analyzed so that the reader may appreciate the breadth of derived contrasts that are possible using the proposed protocol. These analyses were performed for representative subject.

- FLAIR acquisitions offer high T2/PD contrast between lesion and normal tissue; therefore, the conventional analysis methods that have been presented rely on either (1) visual inspection for diagnosis or (2) circumscribing lesions to assess lesion evolution.
- MPRAGE acquisitions are typically exploited for volumetric analyses (Jovicich et al., 2009; Kochunov and Duff Davis, 2010) of structures of interest.
- DTI data are coregistered and fit with a voxelwise tensor model using an automated package and then investigated with tractography or examined for localized changes in contrast based on diffusion anisotropy or isotropy.
- Resting state fMRI analysis includes slice time correction, realignment, and spatial normalization preprocessing steps (Ashburner and Friston, 2005) followed by advanced modeling to reduce physiological noise (Behzadi et al., 2007). Subsequently, functional connectivity is estimated by one of several methods, the two most prominent being temporal correlation with a seed region (Biswal et al., 1995) and spatial ICA (Calhoun et al., 2001; McKeown et al., 1998).
- B₀ Maps are computed from the phase changes between the two acquired datasets (which are complex-valued) and subsequently corrected for phase wraps.
- *B*₁ Maps are computed by coregistering the two acquisition and examining the ratio between the images *S*₂ and *S*₁ (Yarnykh, 2007).
- ASL cerebral blood flow (CBF) can be calculated according to the procedure outlined by van Osch et al. (2009).
- VASO images are CBV-weighted images and can be interpreted as relative CBV maps in the brain as described by Hua et al. (2009a), b and Lu et al. (2005a).
- MT data are analyzed by coregistration and calculation of the magnetization transfer ratio between the MT-prepared and reference acquisitions (Wolff and Balaban, 1989; Smith et al., 2006).
- qT1 data are analyzed by coregistering the two acquired volumes and performing flip angle calculations.
- qT2 data are estimated from a voxel-wise, two-point logarithmic fit of the data.

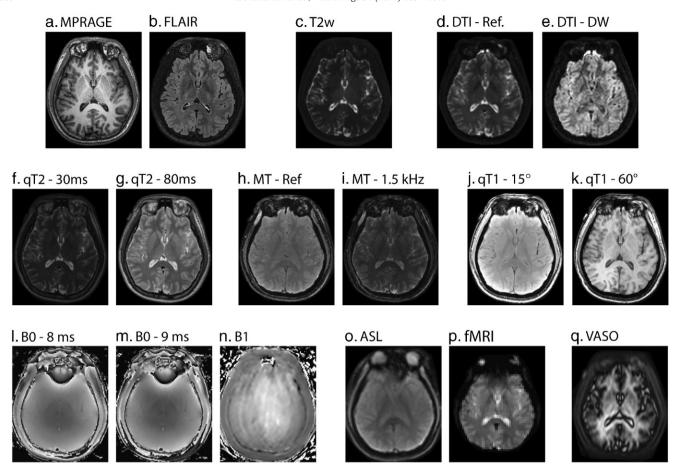


Fig. 1. A single registered slice from the baseline images in the multi-parametric study of a representative subject. Structural scans (a–c) reveal high-resolution information on local tissue classification. Diffusion (d–e), quantitative T2 (f–g), magnetization transfer (h–i), and quantitative T1 (j–k) provide assessments of local micro-structure. Field maps (l–n) enable improved calibration of quantitative findings. Assessment of perfusion (o–p) allows inference on functional characteristics.

Results

Visual inspection of the intra-subject registration of all 535 scans (with a total of 14,562 volumes) verified the absence of gross

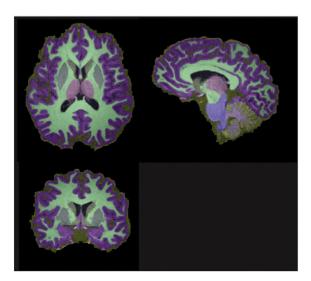


Fig. 2. Representative whole brain segmentation for a single subject. Topologically correct segmentation yields volumetric segmentation of cortical gray matter (purple), cerebral white matter (cyan), thalamus (pink), putamen (slate), caudate (yellow), cerebellar white matter (lavender), brainstem (blue), cerebellar gray matter (marigold), and ventricular CSF (black). Sulcal CSF and dura are labeled separately (tan).

registration errors or uncompensated motion (as illustrated in Fig. 1). Total computation time for registration of all data from 42 sessions (excluding acceleration with parallel processing) was 1 day 20 h 36 min on 2 GHz 64 bit Xeon processors (Intel, Santa Clara, CA). SPECTRE successfully produced individualized skull-stripped volumes for each of the 42 MPRAGE scans, and TOADS segmentation yielded volumetric parcellation of nine cranial structures and a "other/dura"

Table 2 Estimate volume of regions of interest.

Region	Volume (cm³)	Scan-rescan	Intra-class correlation
	$\mu \pm \sigma$	Coefficient of variation	Coefficient (ICC) ^a
Cortical GM	544.4 ± 49.1	2.6%	0.959
Cerebral WM	442.6 ± 45.4	1.5%	0.990
Ventricular CSF	17.2 ± 9.4	4.0%	0.999
Thalamus	19.5 ± 1.4	2.8%	0.912
Putamen	11.8 ± 1.2	3.0%	0.961
Caudate	9.4 ± 1.2	4.0%	0.951
Cerebellar GM	94.9 ± 14.9	4.7%	0.955
Cerebellar WM	30.9 ± 5.0	4.3%	0.961
Brainstem	21.9 ± 2.7	4.7%	0.932
Cranial vault and posterior fossa ^b	1455.1 ± 124.0	2.0%	0.978

Mean volumes are reported for all subjects. Standard deviation is reported across subjects. Scan–rescan CV is reported relative to the mean of the per–subject scan–rescan reproducibility.

All ICCs significantly different from zero at the p<0.001 level.

^b The volume of the cranial vault and posterior fossa is computed as the union of all non-skull structures (identified by SPECTRE).

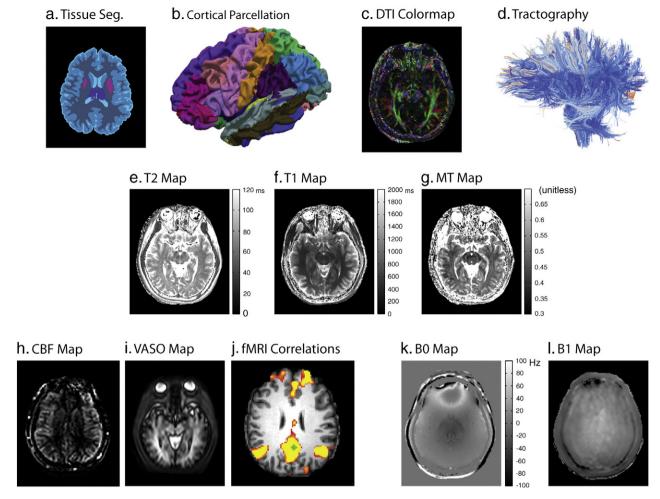


Fig. 3. Illustrative contrasts derived from multi-parametric imaging. Anatomical information, such as tissue classification, cortical parcellation, local tissue diffusion characteristics, and white matter connectivity (a–d) can be inferred from structural and diffusion imaging. Voxel-wise MR properties, such as T1, T2, or MT ratio (e–g), can be inferred from quantitative imaging. Perfusion imaging reveals blood flow characteristics (e.g., perfusion, vascular density, or functional activity, h–j). Resting state fMRI data show the default mode network extracted using precuneus seed (green, MNI coordinates: -2-51 27) correlation. B_0 and B_1 field mapping techniques (k–l) can be used to correct quantitative results and/or spatial distortions.

label classification (as shown in Fig. 2). Table 2 tabulates the volume and reproducibility of each identified structure as estimated by the automated TOADS algorithm. All structures were identified with less

than 5% scan–rescan volume variation (CV < 5%, >0.9 ICC). Fig. 3 illustrates derived contrasts from each of the imaging modalities as analyzed by the methods discussed in the previous section.

Table 3Observed signal: mean intensity by region of interest.

Scan	Cortical GM	Cerebral WM	Ventricular CSF	Thalamus	Putamen	Caudate	Cerebellar GM	Cerebellar WM	Brainstem
	$\mu \pm \sigma$								
FLAIR	1.05 ± 0.01	0.95 ± 0.06	0.47 ± 0.24	0.93 ± 0.04	0.92 ± 0.06	1.06 ± 0.05	1.11 ± 0.05	1.05 ± 0.08	1.04 ± 0.05
MPRAGE (Fast ADNI)	1.02 ± 0.02	1.78 ± 0.14	0.50 ± 0.28	1.82 ± 0.08	1.67 ± 0.06	1.52 ± 0.06	1.09 ± 0.06	1.83 ± 0.16	2.00 ± 0.17
DTI: reference image	1.06 ± 0.02	0.76 ± 0.06	2.66 ± 0.25	0.84 ± 0.06	0.76 ± 0.07	0.89 ± 0.10	1.04 ± 0.18	0.83 ± 0.12	0.90 ± 0.13
DTI: diffusion weighted	0.53 ± 0.03	0.43 ± 0.03	0.34 ± 0.05	0.44 ± 0.03	0.45 ± 0.04	0.50 ± 0.03	0.58 ± 0.10	0.48 ± 0.07	0.45 ± 0.07
DTI geometric reference	1.02 ± 0.02	0.69 ± 0.08	3.77 ± 0.62	1.08 ± 0.09	0.81 ± 0.09	1.09 ± 0.15	0.99 ± 0.07	0.77 ± 0.13	1.00 ± 0.16
Resting state fMRI	1.03 ± 0.04	0.95 ± 0.03	1.93 ± 0.24	1.17 ± 0.05	1.04 ± 0.06	1.16 ± 0.07	1.08 ± 0.12	1.09 ± 0.11	0.99 ± 0.15
B_0 magnitude (TE = 8 ms)	1.11 ± 0.07	1.07 ± 0.07	1.16 ± 0.20	1.47 ± 0.27	1.33 ± 0.27	0.84 ± 0.44	1.48 ± 0.75	1.60 ± 0.64	1.62 ± 0.65
B_0 magnitude (TE = 9 ms)	1.11 ± 0.07	1.07 ± 0.08	1.16 ± 0.19	1.47 ± 0.27	1.31 ± 0.27	0.82 ± 0.45	1.50 ± 0.74	1.62 ± 0.64	1.63 ± 0.66
B_1 magnitude	0.99 ± 0.05	1.08 ± 0.08	0.97 ± 0.23	0.97 ± 0.15	1.08 ± 0.46	0.86 ± 0.31	1.13 ± 0.10	1.10 ± 0.13	0.93 ± 0.12
Arterial spin labeling (ASL)	1.10 ± 0.06	0.97 ± 0.08	1.02 ± 0.22	1.19 ± 0.25	0.94 ± 0.35	0.66 ± 0.25	1.67 ± 0.73	1.61 ± 0.65	1.48 ± 0.57
VASO	1.14 ± 0.08	1.16 ± 0.21	1.45 ± 0.40	1.91 ± 0.49	1.40 ± 0.47	0.99 ± 0.40	2.01 ± 1.35	2.42 ± 1.24	2.12 ± 1.14
qT1 (15°)	1.00 ± 0.01	0.90 ± 0.02	0.78 ± 0.04	0.81 ± 0.02	0.86 ± 0.03	0.87 ± 0.02	0.92 ± 0.12	0.82 ± 0.05	0.76 ± 0.04
qT1 (60°)	1.00 ± 0.01	1.23 ± 0.02	0.68 ± 0.13	1.04 ± 0.04	1.12 ± 0.05	1.02 ± 0.04	1.06 ± 0.10	1.19 ± 0.07	1.11 ± 0.07
qT2 (dual echo: 30 ms)	1.09 ± 0.08	1.06 ± 0.07	1.29 ± 0.20	1.44 ± 0.27	1.31 ± 0.33	1.09 ± 0.35	1.29 ± 0.48	1.40 ± 0.54	1.52 ± 0.46
qT2 (dual echo: 80 ms)	0.65 ± 0.05	0.62 ± 0.05	0.80 ± 0.14	0.90 ± 0.19	0.74 ± 0.23	0.64 ± 0.21	0.78 ± 0.26	0.87 ± 0.36	0.97 ± 0.31
MT: reference	1.09 ± 0.07	0.98 ± 0.05	1.00 ± 0.17	1.21 ± 0.24	1.18 ± 0.34	0.75 ± 0.37	1.35 ± 0.53	1.35 ± 0.44	1.31 ± 0.39
MT: 1.5 kHz	0.62 ± 0.05	0.54 ± 0.03	0.59 ± 0.11	0.70 ± 0.19	0.63 ± 0.21	0.43 ± 0.21	0.75 ± 0.24	0.74 ± 0.24	0.74 ± 0.19

Note σ is the standard deviation of mean intensities across subjects.

 Table 4

 Intra-region homogeneity: coefficient of variation by region of interest.

Scan	Cortical GM	Cerebral WM	Ventricular CSF	Thalamus	Putamen	Caudate	Cerebellar GM	Cerebellar WM	Brainstem
	$\mu \pm \sigma$								
FLAIR	26%±5	16%±3	40% ± 8	$21\% \pm 4$	$14\% \pm 3$	$23\% \pm 7$	$24\% \pm 6$	19%±3	22%±5
MPRAGE (Fast ADNI)	$34\% \pm 7$	$34\% \pm 8$	$37\% \pm 19$	$32\% \pm 5$	$16\% \pm 4$	$24\%\pm10$	$30\% \pm 5$	$27\% \pm 7$	$30\% \pm 15$
DTI: reference image	$33\% \pm 4$	$14\% \pm 4$	$48\% \pm 19$	$26\% \pm 8$	$10\% \pm 3$	$22\% \pm 12$	$34\% \pm 11$	$20\% \pm 9$	$39\% \pm 14$
DTI: diffusion weighted	$16\% \pm 2$	$9\% \pm 1$	$9\% \pm 3$	$9\% \pm 1$	$6\% \pm 2$	$7\% \pm 2$	$17\% \pm 6$	$12\% \pm 3$	$15\% \pm 4$
DTI geometric reference	$42\% \pm 8$	$23\% \pm 8$	$111\% \pm 35$	$50\% \pm 14$	$14\% \pm 4$	$60\% \pm 27$	$36\% \pm 10$	$30\% \pm 16$	$39\% \pm 22$
Resting state fMRI	$30\% \pm 6$	$12\% \pm 2$	$23\% \pm 9$	$21\% \pm 5$	$10\% \pm 4$	$14\% \pm 7$	$27\% \pm 7$	$14\% \pm 3$	$25\% \pm 11$
B_0 magnitude (TE = 8 ms)	$63\% \pm 30$	$61\% \pm 33$	$58\% \pm 35$	$44\% \pm 37$	$39\% \pm 35$	$38\% \pm 22$	$25\% \pm 11$	$22\% \pm 13$	$25\% \pm 9$
B_0 magnitude (TE = 9 ms)	$64\% \pm 30$	$62\% \pm 33$	$59\% \pm 36$	$45\% \pm 38$	$39\% \pm 33$	$37\% \pm 21$	$25\% \pm 11$	$22\% \pm 13$	$25\% \pm 9$
B ₁ magnitude	$54\% \pm 17$	$59\% \pm 25$	$39\% \pm 17$	$27\%\pm14$	$42\% \pm 48$	$41\% \pm 23$	$27\% \pm 14$	$25\% \pm 7$	$22\% \pm 6$
Arterial spin labeling (ASL)	$76\% \pm 36$	$74\% \pm 38$	$60\% \pm 37$	$44\% \pm 44$	$38\% \pm 30$	$38\% \pm 20$	$26\% \pm 9$	$15\% \pm 4$	$15\% \pm 6$
VASO	$98\% \pm 31$	$95\% \pm 29$	$98\% \pm 31$	$106\% \pm 44$	$44\%\pm14$	$52\% \pm 20$	$101\% \pm 52$	$100\% \pm 50$	$101\% \pm 42$
qT1 (15°)	$20\% \pm 2$	$10\% \pm 1$	$5\% \pm 1$	$7\% \pm 1$	$4\% \pm 1$	$5\% \pm 1$	$22\% \pm 9$	$9\% \pm 5$	$14\% \pm 7$
qT1 (60°)	$21\% \pm 3$	$13\% \pm 2$	$24\% \pm 6$	$17\% \pm 4$	$7\% \pm 1$	$14\% \pm 6$	$30\% \pm 13$	$15\% \pm 10$	$24\% \pm 10$
qT2 (dual echo: 30 ms)	$61\% \pm 29$	$60\% \pm 30$	$54\% \pm 23$	$43\% \pm 24$	$43\% \pm 35$	$53\% \pm 24$	$35\% \pm 10$	$37\% \pm 17$	$38\% \pm 11$
qT2 (dual echo: 80 ms)	$45\% \pm 16$	$44\% \pm 15$	$49\% \pm 12$	$47\% \pm 16$	$33\% \pm 23$	$44\% \pm 11$	$36\% \pm 11$	$44\% \pm 24$	$46\% \pm 18$
MT: reference	$65\% \pm 29$	$59\% \pm 32$	$47\% \pm 31$	$34\% \pm 32$	$42\% \pm 37$	$34\% \pm 21$	$27\% \pm 13$	$18\% \pm 12$	$17\% \pm 8$
MT: 1.5 kHz	$41\% \pm 16$	$37\% \pm 17$	$37\% \pm 15$	$34\%\pm21$	$25\% \pm 20$	$25\% \pm 11$	$27\% \pm 8$	$25\% \pm 10$	$24\% \pm 6$

Note σ is the standard deviation of mean intensities across subjects.

Observed image intensities are reported by the MRI scanner in arbitrary units that depend on numerous factors including amplifier gain, multi-channel fusion, conversion to 12- or 16-bit representations, etc. To compensate for this nuisance variability and ensure consistent reporting across modality, Table 3 reports the mean signal intensities normalized by the mean intensity of cerebral gray matter for each subject. Table 4 assesses the intra-region homogeneity for each imaging modality as defined as the standard deviation of the voxels within a region divided by the mean intensity of the corresponding region (i.e., the intra-region coefficient of variation). Table 5 summarizes scanrescan reproducibility measures as the coefficient of variation across repetition, i.e., the standard deviation of the difference of repeated images within a region divided by the mean signal intensity in the region. The scan-rescan reproducibility can be interpreted as the noiseto-signal ratio (i.e., the inverse of the signal-to-noise, SNR); however, this metric is also sensitive to artifacts (systematic variation) and random variation between the scan-rescan events (Landman et al., 2009). Table 6 summarizes scan-rescan reproducibility measures in terms of ICC. Intra-session scan-rescan SNR is reported in Table 7.

Discussion

In typical reproducibility assessments, results are reported for specific contrasts or measures of interest. Here, we take a distinct approach in terms of quantifying the reproducibility of only the directly observed quantities. These characterizations can be particularly informative for SNR, CNR, or power calculations to assess the potential effects of changes in voxel size, slice thickness, number of signal averages, or other simple protocol manipulations. Normal anatomical variability (both temporal and inter-subject characteristics) can make rescan–rescan data complex to interpret. In this study, we focus on isolating variation associated with the acquisition calibration and positioning rather than the anatomical and functional characteristics themselves. Hence, these data could be informative as to the detectable effect sizes over longer time courses.

A primary goal of this study is to measure experimental variability owing to methodology rather than natural biological variability. While fully removing biological variability is impossible, the impact of biological variability on these data can be sufficiently minimized. Actual biological variability is attenuated by having volunteers rescanned within an hour of initial scanning, while detected biological variability is minimized by choosing sufficiently large regions of interest for analysis. The regions of interest used in this study reflect the macroscopic organization of the brain (minimum volume approximately 10 cm³), which are insensitive to small biological differences that may occur between scan and rescan. The observed variability from the ROI primarily reflects (1) tissue intensity level relative to the noise level, (2) propensity of a region to artifacts, and

Table 5Scan-rescan reproducibility: coefficient of variation by region of interest.

Scan	CorticalGM	Cerebral WM	Ventricular CSF	Thalamus	Putamen	Caudate	Cerebellar GM	Cerebellar WM	Brainstem
	CV	CV	CV	CV	CV	CV	CV	CV	CV
FLAIR	6%	3%	26%	5%	3%	8%	4%	2%	3%
MPRAGE (Fast ADNI)	3%	1%	11%	1%	1%	1%	4%	2%	1%
DTI: reference image	6%	3%	5%	5%	2%	6%	13%	12%	13%
DTI: diffusion weighted	5%	2%	7%	2%	2%	2%	13%	10%	8%
DTI geometric reference	7%	6%	7%	7%	3%	9%	7%	5%	6%
Resting state fMRI	6%	2%	5%	4%	2%	3%	8%	3%	6%
B_0 magnitude (TE = 8 ms)	11%	19%	22%	15%	30%	95%	7%	2%	2%
B_0 magnitude (TE = 9 ms)	10%	18%	22%	14%	28%	105%	7%	2%	1%
B ₁ magnitude	10%	13%	6%	6%	9%	10%	2%	2%	2%
Arterial spin labeling (ASL)	14%	28%	67%	27%	95%	161%	6%	2%	2%
VASO	9%	20%	74%	36%	117%	305%	16%	12%	10%
qT1 (15°)	2%	1%	1%	2%	1%	2%	11%	9%	5%
qT1 (60°)	4%	2%	6%	3%	1%	4%	17%	14%	10%
qT2 (dual echo: 30 ms)	14%	22%	24%	18%	28%	72%	6%	5%	4%
qT2 (dual echo: 80 ms)	16%	23%	24%	20%	28%	73%	9%	11%	8%
MT: reference	11%	17%	20%	14%	32%	99%	8%	3%	2%
MT: 1.5 kHz	13%	19%	23%	20%	33%	79%	9%	6%	5%

 Table 6

 Scan-rescan reproducibility: intra-class correlation coefficient (ICC) by region of interest.

Scan	Cortical GM	Cerebral WM	Ventricular CSF	Thalamus	Putamen	Caudate	Cerebellar GM	Cerebellar WM	Brainstem
FLAIR	0.76 ± 0.039	0.77 ± 0.028	0.83 ± 0.032	0.75 ± 0.029	0.70 ± 0.028	0.68 ± 0.057	0.73 ± 0.038	0.75 ± 0.023	0.73 ± 0.028
MPRAGE (Fast ADNI)	0.91 ± 0.014	0.94 ± 0.008	0.88 ± 0.023	0.93 ± 0.006	0.82 ± 0.015	0.86 ± 0.025	0.87 ± 0.019	0.86 ± 0.021	0.82 ± 0.025
DTI: reference image	0.87 ± 0.024	0.88 ± 0.014	0.86 ± 0.020	0.90 ± 0.015	0.82 ± 0.026	0.86 ± 0.025	0.76 ± 0.039	0.76 ± 0.050	0.79 ± 0.053
DTI: diffusion weighted	0.82 ± 0.024	0.82 ± 0.009	0.48 ± 0.024	0.67 ± 0.016	0.51 ± 0.030	0.55 ± 0.033	0.69 ± 0.045	0.69 ± 0.042	0.70 ± 0.038
DTI geometric reference	0.89 ± 0.014	0.87 ± 0.015	0.92 ± 0.014	0.94 ± 0.008	0.80 ± 0.021	0.91 ± 0.019	$\boldsymbol{0.87 \pm 0.020}$	0.84 ± 0.023	0.84 ± 0.019
Resting state fMRI	$\boldsymbol{0.82 \pm 0.048}$	0.72 ± 0.029	$0.63 \pm \text{NaN}$	0.70 ± 0.054	0.62 ± 0.036	$\boldsymbol{0.65 \pm 0.051}$	0.70 ± 0.053	0.65 ± 0.057	0.73 ± 0.044
B_0 magnitude (TE = 8 ms)	0.83 ± 0.026	0.79 ± 0.035	0.83 ± 0.031	0.73 ± 0.051	0.72 ± 0.056	0.75 ± 0.055	0.85 ± 0.029	0.82 ± 0.029	0.78 ± 0.026
B_0 magnitude (TE = 9 ms)	0.79 ± 0.030	$\boldsymbol{0.75 \pm 0.041}$	0.77 ± 0.057	0.74 ± 0.058	0.69 ± 0.071	0.71 ± 0.058	0.80 ± 0.035	0.76 ± 0.040	0.73 ± 0.037
B_1 magnitude	0.48 ± 0.050	0.46 ± 0.061	$\boldsymbol{0.32 \pm 0.070}$	$\boldsymbol{0.33 \pm 0.073}$	0.18 ± 0.061	0.21 ± 0.072	0.63 ± 0.076	0.54 ± 0.070	0.53 ± 0.073
Arterial spin labeling (ASL)	0.92 ± 0.008	0.91 ± 0.010	0.87 ± 0.033	0.80 ± 0.027	0.79 ± 0.044	0.76 ± 0.056	0.81 ± 0.025	0.70 ± 0.031	0.70 ± 0.039
VASO	0.94 ± 0.005	0.93 ± 0.008	0.83 ± 0.034	$\boldsymbol{0.82 \pm 0.041}$	0.82 ± 0.041	0.67 ± 0.056	0.90 ± 0.014	0.88 ± 0.019	0.87 ± 0.016
qT1 (15°)	0.87 ± 0.014	0.89 ± 0.013	0.77 ± 0.024	$\boldsymbol{0.85 \pm 0.020}$	0.61 ± 0.050	0.66 ± 0.053	0.74 ± 0.053	0.74 ± 0.051	0.67 ± 0.072
qT1 (60°)	0.81 ± 0.026	0.79 ± 0.028	0.86 ± 0.023	0.88 ± 0.021	0.63 ± 0.036	0.75 ± 0.061	0.70 ± 0.065	0.71 ± 0.064	0.61 ± 0.066
qT2 (dual echo: 30 ms)	0.92 ± 0.012	0.89 ± 0.014	0.91 ± 0.020	0.86 ± 0.038	0.87 ± 0.029	0.89 ± 0.028	0.87 ± 0.029	0.91 ± 0.027	0.92 ± 0.018
qT2 (dual echo: 80 ms)	0.90 ± 0.016	0.89 ± 0.016	0.91 ± 0.022	0.88 ± 0.039	0.87 ± 0.023	0.89 ± 0.028	0.87 ± 0.030	0.91 ± 0.025	0.93 ± 0.017
MT: reference	0.93 ± 0.007	0.92 ± 0.008	$\boldsymbol{0.88 \pm 0.022}$	$\boldsymbol{0.77 \pm 0.040}$	0.76 ± 0.037	$\boldsymbol{0.80 \pm 0.035}$	0.83 ± 0.031	0.79 ± 0.036	0.79 ± 0.031
MT: 1.5 kHz	0.90 ± 0.011	$\boldsymbol{0.89 \pm 0.013}$	0.91 ± 0.017	0.86 ± 0.030	0.78 ± 0.024	$\boldsymbol{0.82 \pm 0.034}$	$\boldsymbol{0.82 \pm 0.033}$	0.86 ± 0.028	$\boldsymbol{0.89 \pm 0.018}$

Mean ICC shown + standard error.

(3) regional sensitivity to variations in partial-volume effects. These data are therefore intended to provide the background noise level of an experimental methodology that one needs to determine the power of that method in measuring true biological variability within a smaller ROI.

Reproducibility of structural scans was higher than that of images targeting physiologically quantities. In the cortical GM and WM, T1 weighted scans (qT1 and MPRAGE) exhibited the highest reproducibility (mean CV< 2.2%), while DTI, fMRI, and FLAIR exhibited good reproducibility (mean CV~ 5%). MT and quantitative T2 exhibited mean cortical reproducibility of CV 17%. ASL was the least reproducible in the cortical GM and WM with a mean CV of 21%. Notably, the ASL intensity in white matter exhibited a CV of 28%. Note that ASL is a perfusion sensitized measurement and WM has very low perfusion relative to cortical GM (which had a CV of 14%). VASO reproducibility was also low with a CV of 15%. Increased variability for quantitative scans is to be expected both because of physiological variability as and the increased sensitivity for artifacts.

Although the structural scans (MPRAGE and FLAIR) exhibited consistent reliability between the brain and cerebellum, reproducibility of the sequences was not uniformly consistent for all scan types. ASL was exceptionally stable for cerebellar and brainstem structures (CV of <3%) while quantitative T2, MT, and field mapping approaches were also improved compare with cerebral structure (CV reduced by >11%). Yet, the quantitative T1 exhibited much lower reproducibility (CV increased by <7%) for brainstem regions. For deep brain structures (thalamus,

Table 7 Intra-session SNR measures.

Scan	Corpus callosum	Peripheral WM	Globus palidus
FLAIR	10.9	17.0	13.4
MPRAGE (Fast ADNI)	25.6	30.2	25.5
DTI: reference Image	13.1	36.7	30.1
DTI: diffusion weighted	8.2	11.8	11.0
DTI geometric reference	5.2	15.2	16.7
Resting state fMRI	24.2	36.8	40.6
B_0 magnitude (TE = 8 ms)	13.9	60.6	65.1
B_0 magnitude (TE = 9 ms)	6.9	55.8	39.2
B_1 magnitude	66.4	80.7	69.1
Arterial spin labeling (ASL)	48.8	56.5	79.3
VASO	26.3	68.6	30.4
qT1 (15°)	25.2	48.0	52.4
qT1 (60°)	25.9	46.6	35.1
qT2 (dual echo: 30 ms)	24.3	51.9	49.8
qT2 (dual echo: 80 ms)	11.7	30.3	23.6
MT: reference	25.9	44.4	38.3
MT: 1.5 kHz	26.2	42.2	29.5

putamen, caudate), there was strong correlation (ρ =0.74) between acquired slice thickness and CV of these structure. For deep brain structures, T1 weighted studies had high average reproducibility (<3% CV) while the ASL CV neared unity (94%) and the mean CV with VASO was 1.53. MT and quantitative T2 also performed poorly in the caudate (80% CV). CSF exhibited relatively low reproducibility (>5% CV) for all scans except quantitative T1, in which it was consistently of moderate intensity (~70% that of GM). The impact variability can be appreciated from the increased variability on bordering structures, especially those in which CSF is bright. For example, the caudate was poorly reproduced in scans with T2 weighted contrasts.

Herein, we tabulate the reproducibility of image intensities for large-scale neurological structures. These reproducibility measures are, by construction, less than that which one would expect if smaller, more homogeneous, regions were carefully placed. Specifically, we have intentionally included the effects of scan-rescan variation in artifact, distortion, partial voluming, and non-rigid motion that might corrupt subsequent analysis. The regions of interest were chosen to be representative of the large-scale regions that (1) are of general interest to community and (2) are reproducible with fully automated methods. More detailed characterizations would be valuable to specific fields, such as inclusion of a cortical parcellation or more precise characterization of white matter. However, these approaches would introduce additional methodological dependence and would likely benefit from careful consideration of the specific problem domain. Use of these data for comparative evaluation of anatomical characterizations (and their associated MR-derived contrasts) would be an interesting area of future research. Alternatively, regions of interest could be placed as to avoid these potential confounds if these effects would not be interfere with the data interpretation. We note that the variability measures themselves (e.g., resting state fMRI and diffusion weighted MRI intensities) are not always of direct interest. Rather, the scan-rescan variability measured presented herein can be used to model (either empirically or in simulation) the variability of derived parameters of interest. These data have been made public so that study-specific local regions of interest may be readily compared to the summary measures presented.

A direct analysis of the reliability and reproducibility does not mitigate the information contained in the resulting images. For example, ASL has the lowest reproducibility, but it is the sole method for studying cerebral perfusion in the absence of contrast injection, so it is invaluable, although maybe not as reproducible as an approach that measures large-scale changes in tissue water. Without a doubt, SNR plays a clear role in the reproducibility specific, yet a precise assessment of SNR for each contrast would necessitate either additional data (i.e., scan-rescan information within a session or

noise-only scans) or making assumptions as to the where one could isolate of signal regions with true homogenous signal or use parametric modeling assumptions. As presented, this multi-parametric protocol can serve as a baseline set of sequences to which new generations of acquisition techniques can be compared.

Given the rather limited sample size (n=21), it is difficult to control for age and gender effects, which are known to modulate both brain structure and function. This heterogeneity does not necessarily undermine the assessment of within subject variation (so long as variability is not particularly associated with healthy aging). Furthermore, for power analyses, a hypothesis on effect size is also required. Here, we leave it to the reader to estimate effect size for the analyses methods and contrasts relevant to their endeavors. If imaging variability is believed to be strongly associated with demographics for a particular condition, the data are fully available so that cohorts may be repartitioned into representative populations and appropriate estimates of imaging characteristics may be estimated. The protocol has been fully released so additional acquisitions on select target populations with equivalent sequences are also possible and may be compared with those that have been presented.

There are ample opportunities to use this dataset to assess of specific metrics (e.g., quantitative parameters) and/or multi-parametric mapping (e.g., functional connectivity measures on cortical surfaces labeled with diffusion inferred connectivity). In many cases, the numerous potential approaches to derive the same metric are available and it would be an interesting scientific effort to compare such methods. Furthermore, these best practice methods may vary by intended application and require substantial exposition to detail the precise variant of a method used during analysis. Although these analyses are fascinating avenues for continuing study, such efforts are beyond the scope of this work. The authors are actively pursuing characterization of several approaches; however, independent efforts by other groups are also encouraged.

Conclusion

All resources for this article have been publicly shared through the resource website of the national resource for quantitative functional MRI (http://mri.kennedykrieger.org/databases.html) and the "multimodal" project on the NITRC portal (http://www.nitrc.org/projects/multimodal/). The community is encouraged to use these data for investigation of the topics relevant to their research. Investigators who accept the BIRN Repository Data License may access, redistribute, and publish the data without justification of intended use. Forums are provided for researchers who, at their discretion, may seek to coordinate or collaborate on investigations.

In conclusion, this effort provides the neuroimaging community with a data resource that may be used for development, optimization, and evaluation of algorithms that exploit the diversity of modern MRI modalities. The data may contribute to or serve as *a priori* information for study planning, effect size estimation, or as normative reference data from volunteers. Alternatively, the data may be used within an education or tutorial setting and hence provide realistic examples of the data available within clinical research. By optimizing a collection of modern MRI sequences to mutually run in under an hour, we have created a baseline for planning and more specialized study design at 3 T.

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